

STUDY ON PHYTIC ACID CONTENT IN SOME ROMANIAN CEREAL SEEDS

MARCEL AVRAMIUC^{1*}

Received: 26th of April 2018 / Revised: 10th of May 2018

Accepted: 27th of May 2018 / Published: 29th of June 2018

Keywords: phytic acid, cereal, flour, bran

Abstract. The phytic acid content of the flour and bran from seeds, belonging to four cereal species, as well as the effect of thermal treatment on this acid was the purpose of this paper. The biological material was represented by dried seeds (moisture content 10%) of twelve Romanian cultivars (c), belonging to four cereal species: wheat (3 c), rye (3 c), oat (3 c) and maize (3 c). The seeds of each species were ground to get integral flour and bran. The phytic acid content was analyzed, using Garcia-Villanova method, in the flour and bran, before and after thermal processing (180°C for 50 minutes). They also determined pH values of the integral flour and bran in the four cereal species. Analyzing the phytic acid in integral flour and bran of raw samples, the highest and close values were in rye and maize, and the least ones in wheat and oat. Thermal processing of the both products (flour and bran) led to highest and significant reducing percent of phytic acid in rye, followed by oat and wheat samples with close values, the least reducing percent of phytic acid being in maize samples.

INTRODUCTION

Phytic acid (myo-inositol hexakisphosphate, InsP6) is an organic compound present in plant seeds and grains (Rose, 1912; Averill and King, 1926; Harland and Oberleas, 1987; Reddy et al., 1989), roots and tubers (Harland and Oberleas, 1987; Wolters et al., 1993; Ravindran et al., 1994), fruits and vegetables (Harland and Oberleas, 1987; Wolters et al., 1993; Ravindran et al., 1994), nuts (Harland and Oberleas, 1987; Wolters et al., 1993, cited by Reddy, 2002) and in other vegetable sources.

Phytic acid is a strong chelator of divalent minerals such as copper, calcium, magnesium, zinc, and iron (Sathe and Reddy, 2002), and studies on animals and humans highlighted that phytates decrease mineral bioavailability by forming complexes with these minerals (Reddy et al., 1989; Krebs, 2000).

According to some authors (Cheryan, 1980; Thompson, 1993) phytic acid can also form complexes with proteins as phytate-protein and phytate-mineral-protein, adversely influencing protein digestion and bioavailability. With starch, phytate can bind either directly, by hydrogen bonding, with a phosphate group or indirectly, through the proteins (Rickard and Thompson, 1997), which may result in a decrease of starch solubility and digestibility (Sathe and Reddy, 2002).

Food phytates have not only negative effects, as mentioned before, but also beneficial effects including lowering of serum cholesterol and triglycerides, and protection against certain diseases such as cardiovascular diseases, renal stone formation, and certain types of cancers (Thompson, 1993; Rickard and Thompson, 1997; Zhou and Erdman, 1995; Shamsuddin et al., 1997; Grases et al., 2000; Urbano et al., 2000; Graf, 1983, cited by Sathe and Reddy, 2002).

In this paper, the phytic acid content of the flour and bran from seeds, belonging to four cereal species, was studied, as well as the effect of the heat treatment on this compound.

MATERIALS AND METHODS

Research materials. The biological material, coming from the last year crop, was represented by seeds of twelve romanian cultivars (c) belonging to four cereal species: wheat (3 c), rye (3 c), oat (3 c) and maize (3 c). The seeds were dried (moisture content 10%), and after that they were ground to get integral flour and bran, for each species.

Procedure and research methods. The phytic acid content was analyzed in integral flour and bran, before and after the thermal processing of the same materials, in oven at 180°C for 50 minutes. They also determined pH values of the integral flour and bran in the four cereal species.

The phytic acid was determined using Garcia-Villanova method (Bordei et al., 2007). They weighed 2 g of flour or bran, added 40 ml of HCl-Na₂SO₄ solution and leaved to stand for 90 minutes, stirring intermittently (to extract phytic acid). They filtered and collected 25 ml of the filtrate, which were placed in a stoppered tube, then added 20 ml of HCl-Na₂SO₄ solution, 20 ml hydrochloride solution of ferric chloride, and 20 ml of 20% sulfosalicylic acid solution. It boiled water in the closed tube for 15 minutes on a water bath, then cooled under a stream of water. It formed the ferric filtrate

precipitate which was separated by filtration. From the clear (obtained) filtrate, it took 25 ml, placed in an Erlenmeyer flask, added about 175 ml of distilled water, and then adjusted the pH to 2.5 ± 0.5 with glycine. The mixture obtained was heated to 70°C and was titrated the excess of Fe^{3+} (which has not precipitated with phytic acid), with 0.15M EDTA solution until the color shifted to bright yellow. The result was expressed in g phytic acid per 100 g product (%). Four replicates for each determination were used to get the data of experiments.

Determination of pH was made with a digital pH meter supplied by Hanna Instruments.

Statistical analysis. The experimental data were statistically processed using SAS Version 8.02 (SAS Institute, 2005). To analyze the significance of differences among samples, generalized linear model analysis was carried out, and for multiple comparisons, Duncan's multiple range test ($P < 0.05$) was used.

RESULTS AND DISCUSSIONS

In the Table 1 are rendered the values of phytic acid and pH in samples coming from four cereal species, before and after thermal processing.

Table 1. Phytic acid content mean values ($\pm$ SD) and pH in flour and bran of cereal seeds

Cereal samples	Raw samples				Thermal processed samples	
	Flour		Bran		Flour	Bran
	pH	Phytic acid (%)	pH	Phytic acid (%)	Phytic acid (%)	Phytic acid (%)
W1	5.3	1.22 $\pm$ 0.5FG*	5.2	3.17 $\pm$ 0.9AB	0.96 $\pm$ 0.03GH	2.54 $\pm$ 0.7C
W2	5.4	1.16 $\pm$ 0.7FG*	5.3	2.98 $\pm$ 0.4AB	0.89 $\pm$ 0.08H	2.42 $\pm$ 0.8CD
W3	5.2	1.07 $\pm$ 0.6G	5.3	2.83 $\pm$ 0.5B	0.83 $\pm$ 0.05H	2.33 $\pm$ 0.5D
R1	4.6	1.20 $\pm$ 0.8FG	4.7	3.17 $\pm$ 0.5AB	0.81 $\pm$ 0.04H	2.27 $\pm$ 0.4D
R2	4.7	1.45 $\pm$ 0.4F	4.7	3.34 $\pm$ 0.4A	0.92 $\pm$ 0.05GH	2.34 $\pm$ 0.6D
R3	4.6	1.27 $\pm$ 0.7FG	4.8	3.28 $\pm$ 0.7A	0.81 $\pm$ 0.07H	2.35 $\pm$ 0.3D
O1	5.4	1.08 $\pm$ 0.6G	5.2	3.10 $\pm$ 0.5AB	0.82 $\pm$ 0.06H	2.48 $\pm$ 0.5CD
O2	5.4	1.12 $\pm$ 0.9G	5.3	2.78 $\pm$ 0.8BC	0.83 $\pm$ 0.04H	2.27 $\pm$ 0.8D
O3	5.3	1.18 $\pm$ 0.7FG	5.3	3.04 $\pm$ 0.6AB	0.91 $\pm$ 0.02GH	2.40 $\pm$ 0.9CD
M1	5.8	1.17 $\pm$ 0.4FG	5.7	3.10 $\pm$ 0.3AB	0.97 $\pm$ 0.06GH	2.62 $\pm$ 0.6C
M2	5.8	1.31 $\pm$ 0.5F	5.5	3.29 $\pm$ 0.7A	1.07 $\pm$ 0.8G	2.80 $\pm$ 0.5BC
M3	5.7	1.25 $\pm$ 0.2FG	5.6	3.24 $\pm$ 0.6A	1.03 $\pm$ 0.05G	2.72 $\pm$ 0.2BC

SD=standard deviation; W1-3=wheat cultivars; R1-3=rye cultivars; O1-3=oat cultivars; M1-3=maize cultivars;

*Means with the same letters within a column are not statistically different ($P < 0.05$)

As seen from Tab. 1, the pH values of cereals flour ranged between 4.6 (rye) and 5.8 (maize), and pH of cereals bran between 4.7 (rye) and 5.7 (maize).

Analyzing the phytic acid in integral flour of raw (unprocessed) samples, the highest values were in rye (1.45%) and maize (1.31%) with close values, and the least ones in wheat (1.07%) and oat (1.08%) with close values too ($P < 0.05$).

According to some authors (Lolas *et al.*, 1976; Eechkhout and Depaepe, 1994; DeBoland *et al.*, 1975; Kikunaga *et al.*, 1985; Harland and Prosky, 1979), cited by Reddy N.R. (2002), the phytic acid content of some cereal grains ranges between the following limits: 0.39%-1.35% (wheat), 0.54%-1.46% (rye), 0.42%-1.16% (oat) and 0.75%-2.22% (maize).

In the bran of raw samples, the phytic acid registered the highest and close values in rye (3.28% - 3.34%) and maize (3.24% - 3.29%), and the least one in oat (2.78%) and wheat (2.83%).

As seen, the phytic acid content in bran was higher then in flours, because, according to Reddy (2002), in most cereals phytate is concentrated in the germ and aleurone layers (pericarp)

of the grain cells. In maize 88% of phytate is concentrated in the germ portion of the grain (O'Dell *et al.*, 1972).

By some data, the phytic acid content of wheat bran ranges between 2.02% and 5.27% (Harland and Prosky, 1979; Ellis and Morris, 1982; Harland and Oberleas, 1986; Kirby and Nelson, 1988; Lehrfeld and Wu, 1991; Eechkhout and Depaepe, 1994; Gualberto *et al.*, 1997; Kasim and Edwards, 1998) and the phytic acid of oat bran between 0.60% and 1.42% (Frolich and Nyman, 1988; Gualbert *et al.*, 1997; Kasim and Edwards, 1998).

After thermal processing, the phytic acid of flour registered the highest value in maize (1.07%), and the least ones in rye (0.81%), oat (0.82%) and wheat (0.83%) with close values ($P < 0.05$).

The highest phytic acid content of thermal processed bran was in maize (2.80%) and the least ones in oat, rye and wheat samples, with close values (2.27% - 2.33%).

Analyzing the average values of phytic acid in flour and bran it can be observed that in all cases they were reduced by thermal processing.

The table 2 highlights the reduction percentage of phytic acid in cereals flour and bran due to thermal processing.

Table 2. Reduction percentage of phytic acid average values in flour and bran thermal processed, compared to raw

Cereal samples	Phytic acid reduction percents	
	Flour	Bran
W1	-21.31% d	-19.87% e
W2	-23.27% cd*	-18.79% e
W3	-22.43% cd*	-17.66% ef
R1	-32.50% ab	-28.39% bc
R2	-36.55% a	-29.94% b
R3	-36.22% a	-28.35% bc
O1	-24.07% cd	-20.00% e
O2	-25.89% c	-18.34% ef
O3	-22.88% cd	-21.05% de
M1	-17.09% f	-15.48% fg
M2	-18.32% e	-14.89% fg
M3	-17.60% ef	-16.05% fg

W1-3=wheat cultivars; R1-3=rye cultivars; O1-3=oat cultivars; M1-3=maize cultivars; *Means with the same letters within a column are not statistically different ($P < 0.05$)

Thus, compared to raw samples, the thermal processing of flour, at 180°C for 50 minutes, led to highest and significant reducing percent of phytic acid in rye (-36.55%), followed by oat and wheat samples with close values; the least reducing percent of phytic acid (-17.09%) was in maize samples ($P < 0.05$).

Under the same conditions, the thermal processing of bran led to highest and significant reducing percent of phytic acid in rye (-29.94%), followed by oat and wheat samples with close values, the least reducing percent (-14.89%) being also in maize samples ($P < 0.05$).

One can observe that thermal processing, at 180°C for 50 minutes, caused a higher reducing of phytic acid content in flour, compared to bran. It is possible that the presence of cellulose and lignin in the bran composition to partially protect the phytic acid from the harmful

action of high temperatures.

Some of these results are consistent with observations and data reported by certain authors.

A significant percent of phytate hydrolysis can take place due to activation of endogenous phytases and acid phosphatases, during the early part of the cooking phase (Sathe and Venkatachalam, 2002).

According to some authors (Reddy *et al.*, 1989; McKenzieparnell and Davies, 1986; Sathe and Venkatachalam, 2002), the phytic acid content of cereals is reduced by fermentation and bread making with 8.9% in the whole-meal breads (unleavened) and with 66-100% in white breads.

It seems that pH and cooking temperature are important factors reducing the content of phytic acid, because in doughs with pH 4.3–4.6 phytic acid is more effectively reduced than in doughs with higher pH (Fretzdorff and Brümmer, 1992).

In our experiment the least pH values (4.6–4.7) were registered in rye samples, and also at these samples were recorded the highest reductions in the content of phytic acid by exposure to high temperature.

CONCLUSIONS

The analyze of phytic acid content in the integral flour and bran of grains, belonging to four cereal species, evidenced some significant differences between species.

Thus, the highest phytic acid content of integral flour and bran was in rye and maize with close values, and the least one in wheat and oat with close values too.

In all analyzed samples, the phytic acid content of bran was significantly higher than of integral flours.

The thermal processing of flour and bran (180°C for 50 minutes) led to highest and significant reducing percent of phytic acid in rye, followed by oat and wheat samples with close values, the least reducing percent of phytic acid being in maize samples.

The thermal processing caused a higher reducing of phytic acid content in flour, compared to bran, probable due to the presence of cellulose and lignin in the bran composition, which partially protect this acid from the harmful action of high temperatures.

REFERENCES

- Averill H.P., King C.G. (1926) - *The phytin content of foodstuffs*. J. Amer. Chem. Soc. **48**:724-728
- Bordei D. (coord.), Bahrim G., Pâslaru V., Gasparotti C., Elisei A., Banu I., Ionescu L., Codină G. (2007) - *Quality control in the bakery industry. Analytical methods*. Ed. Academica, Galați, 236-237
- Cheryan, M. (1980) - *Phytic acid interactions in food systems*. CRC Crit. Rev. Food Sci. Nutr. **13**:297–325
- DeBoland A., Garner G.B., O'Dell B.L. (1975) - *Identification and properties of phytate in cereal grains and oilseed products*. J. Agric. Food Chem. **23**:1186–1189.
- Eechkhout W., Depaepe M. (1994) - *Total phosphorus, phytate phosphorus, and phytase activity in plant feed stuffs*. Animal Feed Sci. Technol. **47**:19–29
- Ellis R., Morris E.R. (1982) - *Comparison of ion-exchange and iron-precipitation methods for analysis of phytate*. Cereal Chem. **59**:232–233.
- Fretzdorff B., Brümmer J.-M. (1992) - *Reduction of Phytic Acid During Breadmaking of Whole-Meal Breads*. Cereal Chem. **69**, 266–270
- Frolich W., Nyman M. (1988) - *Minerals, phytate, and dietary fibre in different fractions of oat grain*. J. Cereal Sci. **7**:73–80
- Graf E. (1983) - *Applications of phytic acid*. J. Amer. Oil Chem. Soc. **60**:1861–1867
- Grases F., March J.G., Prieto R.M., Simonet B.M., Costa-Bauza A., Garcia-Raja A., Conte A. (2000) - *Urinary phytate in calcium oxalate stone formers and healthy people. Dietary effects on phytate excretion*. Scan. J. Urology Nephrology **34**:162–164.

- Gualberto D.G., Bergman C.J. Kazemzadeh M., Weber C.W. (1997) – *Effect of extrusion processing on the soluble and insoluble fiber and phytic acid contents of cereal brans*. Plant Foods Hum. Nutr. **51**:187–198.
- Harland B.F., Prosky L., (1979) - *Development of dietary fiber values for foods*. Cereal Foods World **24**:390–394.
- Harland B.F., Oberleas D. (1986) - *Anion-exchange method for determination of phytate on foods: Collaborative study*. J. Assoc. Off. Anal. Chem. **69**:667–670
- Harland B.F., Oberleas D. (1987) - *Phytate in foods*. World Rev. Nutr. Diet. **52**:235–259
- Kasim A.B., Edwards H.M. (1998) - *The analysis of inositol phosphate forms in feed ingredients*. J. Sci. Food Agric. **76**:1–9
- Kikunaga S., Takahashi M., Huzisige H. (1985) - *Accurate and simple measurement of phytic acid contents in cereal grains*. Plant Cell Physiol. **26**:1323–1330
- Kirby L.K., Nelson T.S. (1988) - *Total and phytate phosphorus content of some feed ingredients derived from grains*. Nutr. Rep. Intern. **137**:277–280
- Krebs N.F. (2000) - *Overview of zinc absorption and excretion in the human gastrointestinal tract*. J. Nutr. **130**:1374S–1377S
- Lehrfeld J., Wu Y.V. (1991) - *Distribution of phytic acid in milled fractions of Scout 66 hard red winter wheat*. J. Agric. Food Chem. **39**:1820–1824
- Lolas G.M., Palamids N., Markakis, P. (1976) -. *The phytic acid, total phosphorus relationship in barley, oats, soybeans, and wheat*. Cereal Chem. **53**:867–871
- Mckenziepamell J.M., Davies N.T. (1986) - *Destruction of phytic acid during home breadmaking*. Food Chem. **22**:181–192
- O'Dell B.L., DeBoland A., Koirtiyohann R. (1972) - *Distribution of phytate and nutritionally important elements among the morphological components of cereal grains*. J.Agric.Food Chem. **20**:718–721
- Ravindran V., Ravindran G., Sivalogan S (1994) - *Total and phytate phosphorus contents of various foods and feedstuffs of plant origin*. Food Chem. **50**:133-136
- Reddy N.R., Pierson M.D., Sathe S.K., Salunkhe D.K. (1989) - *Phytates in Cereals and Legumes*. CRC Press Inc., Boca Raton, Florida
- Reddy N.R. (2002) - *Occurrence, Distribution, Content, and Dietary Intake of Phytate*, in: Food Phytates, ed. by Rukma N. Reddy, Shridhar K. Sathe, CRC Press LLC, 2000 N.W. Corporate Blvd., Boca Raton, Florida 33431
- Rickard S.E., Thompson L.U. (1997) -. *Interactions and biological effects of phytic acid*. In Antinutrients and Phytochemicals in Food, Shahidi, F. (Ed.), ACS Symposium Series 662, American Chemical Society, Washington, DC, 294–312
- Rose A.R. (1912) - *A resum of the literature on inosite phosphoric acid with special reference to the relation of that substance to plants*. Biochem. Bull. **2**:21-49
- Shamsuddin A.M., Vucenik I., Cole K.E. (1997) - *IP6: A novel anti-cancer agent*. Life Sci. **61**:343–354
- SAS Institute (2005) - *SAS User's Guide*. Statistical Analysis System Institute, Cary, NC
- Sathe S.K., Reddy N.R. (2002) - *Introduction*, in: Food Phytates, ed. by Rukma N. Reddy, Shridhar K. Sathe, CRC Press LLC, 2000 N.W. Corporate Blvd., Boca Raton, Florida 33431
- Sathe S.K., Venkatachalam M. (2002) - *Influence of Processing Technologies on Phytate and Its Removal* in: Food Phytates, ed. by Rukma Reddy, Shridhar K. Sathe, CRC Press LLC, 2000 N.W. Corporate Blvd., Boca Raton, Florida 33431
- Thompson L.U. (1993) - *Potential health benefits and problems associated with antinutrients in foods*. Food Res. Intern. **26**:131–149
- Urbano G., Lopez-Jurado M., Aranda P., Vidai-Valverde C., Tenorio E., Porres J. (2000) - *The role of phytic acid in legumes: antinutritional or beneficial function?* J. Physiol. Biochem. **56**:283–294.
- Wolters M.G.E., Diepenmaat H.B., Hermus R.J.J., Voragen A.G.J. (1993) - *Relation between in vitro availability of minerals and food composition: A mathematical model*. J. Food Sci. **58**:1349-1355.
- Zhou J.R., Erdman J.W. (1995) - *Phytic acid in health and disease*. Crit. Rev. Food Sci. Nutr. **35**:495–508.

ACKNOWLEDGMENTS

Many thanks to Suceava Genebank and to Suceava Agricultural Research and Development Station for the biological material provided.

¹ Faculty of Food Engineering, Stefan cel Mare University of Suceava

* avramiucm@fia.usv.ro

